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# Current Journal of Applied Science and Technology



## **Editorial Policy**

#### Journal Information:

New name: Current Journal of Applied Science and Technology ISSN: 2457-1024 Past name: British Journal of Applied Science & Technology ISSN: 2231-0843 (old) NLM ID: 101664541 Frequency: 15 Days Journal DOI: http://dx.doi.org/10.9734/cjast (http://dx.doi.org/10.9734/cjast) Peer-review model: Advanced OPEN peer review NAAS Score (2019): 5.32 (JrnID: B148) Digital Archiving: Journal Repository (JR), LOCKSS, CLOCKSS

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We have migrated to transparent and toughest 'Advanced OPEN peer review' system (Detailed general information is available in this link). High-quality manuscripts are peer-reviewed by minimum two peers in the same field. OPEN peer review system provides the provision to reveal the identities of the authors and reviewers to each other during the review process. In order to add transparency further, details of all reviewers and academic editors are published in the first page of every published paper (in the Article Information section: see example). As a final step to provide highest level transparency in the process, all review comments, authors' feedbacks, all versions of the manuscript and editorial comments are published (along with date) with the paper in 'Review History' link (See example 1, example 2, example 3, etc). This transparent process will help to eradicate any possible malicious/purposeful interference by any person (publishing staff, reviewer, editor, author, etc) during peer review. As a result of this unique system, all reviewers will get their due recognition and respect, once their names are published in the papers (Example Link). If reviewers do not want to reveal their identities, we will honour that request. In that case, only the review reports will be published as 'anonymous reviewer report'.

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We strongly discourage any attempt by the authors to contact the reviewer directly to influence the review process. We also strongly discourage any attempt by the reviewers to contact the authors directly. General guidelines for Peer-review Process are available below.

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It is a prerequisite to submit, with the manuscript, the names, addresses and e-mail addresses of 4 potential reviewers (When suggesting peer reviewers, please follow these guidelines to avoid any probable conflict of interest. Suggested reviewers should not: i) be from the same department or division as one of the authors (the same university, state, country should also be avoided); ii) have been a research guide or student of one of the authors within the past 10 years; iii) have collaborated with one of the authors within the past 10 years; iv) be employees of non-academic organizations with which one of the authors has collaborated within the past 10 years). It is the sole right of the editorial team to decide whether suggested reviewers to be used or not.

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2. Users, who want to comment, are encouraged to register on website. But if anybody doesn't want to register, we'll respect the decision. In order to honour 'free flow of thoughts' unregistered user are also welcome to comment. Social login is also encouraged.

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Plagiarism policy of this journal is mainly inspired by the plagiarism policy of The Nature. Plagiarism policy of this journal is described below:

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2. Plagiarism can be said to have clearly occurred when large chunks of text have been cut-and-pasted. Such manuscripts would not be considered for publication in this journal. Papers with confirmed plagiarisms are rejected immediately.

3. But minor plagiarism without dishonest intent is relatively frequent, for example, when an author reuses parts of an introduction from an earlier paper.

4. Duplicate publication, sometimes called self-plagiarism, occurs when an author reuses substantial parts of his or her own published work without providing the appropriate references. This can range from getting an identical paper published in multiple journals, to 'salami-slicing', where authors add small amounts of new data to a previous paper. Self-plagiarism, also referred to as 'text recycling', is a topical issue and is currently generating much discussion among editors. Opinions are divided as to how much text overlap with an author's own previous publications is acceptable. We normally follow the guidelines given in COPE website. Editors, reviewers, and authors are also requested to strictly follow this excellent guideline (Reference: Text Recycling Guidelines: http://publicationethics.org/text-recycling-guidelines).

5. In case of 'suspected minor plagiarism', authors are contacted for clarification. Depending on all these reports, reviewers and editors decide final fate of the manuscript. If the manuscript is finally accepted and published, then to maintain transparency, all these reports are published in 'publication history' of the paper by following Advanced OPEN peer review system. The journal editors judge any case of which they become aware (either by their own knowledge of and reading about the literature, or when alerted by referees) on its own merits.

6. Use of the automated software is helpful to detect the 'copy-paste' problem. All submitted manuscripts are checked by the help of different databases, eTBLAST, Plagiarism Detection tools, etc. At the same time scientific implication of the case ('suspected minor plagiarism'), also judged by reviewers and editors. Plagiarism Detection tools are useful, but they should be used in tandem with human judgment and discretion for the final conclusion. Therefore, suspected cases of plagiarisms are judged by editors on 'case-to-case basis'.

7. Editors have the final decision power for these cases.

Some useful information is available at the following links:

- 1. http://www3.imperial.ac.uk/library/researchers/plagiarismdetection
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Publisher Ms. M. B. Mondal, Ph.D. Email: director@sciencedomain.org

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## **Editorial Team**

### **Chief Editors**

Dr. Harry E. Ruda

Professor,

Stan Meek Chair Professor in Nanotechnology,

University of Toronto, Director, Centre for Advanced Nanotechnology,

University of Toronto, Canada

Email: harry.ruda@utoronto.ca, ecanmpm@ecf.utoronto.ca

Profile link: http://mse.utoronto.ca/faculty-staff/professors/ruda/ (http://mse.utoronto.ca/faculty-staff/professors/ruda/)

#### Dr. Singiresu S. Rao

Professor,

Department of Mechanical and Aerospace Engineering,

University of Miami, Coral Gables, USA

Email: srao@miami.edu, srao@umail.miami.edu

Profile link: https://miami.pure.elsevier.com/en/persons/singiresu-s-rao (https://miami.pure.elsevier.com/en/persons/singiresu-s-rao)

Short Biosketch (http://www.journalcjast.com/index.php/CJAST/singiresu-s-rao)

#### Dr. Meng Ma

Associate Professor, Anhui University, Hefei, Anhui, China And Icahn Institute for Genomics and Multiscale Biology, Icahn School of Medicine at Mount Sinai, New York, USA Email: meng.ma@mssm.edu, mengma2@gmail.com, mameng@mail.ustc.edu.cn

Profile link: NA

<u>Short Biosketch</u> (http://www.journalcjast.com/index.php/CJAST/meng-ma)

## Academic Editors

#### Dr. Nan Wu

Associate Professor,

Department of Mechanical Engineering, University of Manitoba, Winnipeg, Canada Email: nan.wu@umanitoba.ca Profile link: https://goo.gl/QgaaPD (https://goo.gl/QgaaPD)

#### Dr. Rodolfo Dufo Lopez

Professor,

Department of Electrical Engineering, University of Zaragoza, Spain

Email: rdufo@unizar.es

Profile link: NA

#### Dr. Diyuan Li

Professor,

School of Resources and Safety Engineering, Central South University, China

Email: diyuan.li@csu.edu.cn

Profile link: NA

#### Dr. Chien-Jen Wang

Professor,

Department of Electrical Engineering, National University of Tainan, Taiwan

Email: cjwang@mail.nutn.edu.tw

Profile link: NA

#### Dr. Teresa De Pilli

Assistant Professor, Department of Science of Agriculture of Food of Environment (SAFE), University of Foggia, Via Napoli, Italy Email: t.depilli@unifg.it, teresadepilli@tiscali.it Profile link: NA

#### Dr. Jerzy Nowacki

Professor,

Faculty of Mechanical Engineering And Mechatronics

West Pomeranian University of Technology, Szczecin, Poland Email: jnowacki@zut.edu.pl Profile link: NA

#### Dr. Elena Lanchares Sancho

Associate Professor, Department of Mechanical Engineering, University of Zaragoza, Zaragoza, Spain Email: elanchares@unizar.es, elanchar@unizar.es Profile link: http://amb.unizar.es/people/elena-lanchares/ (http://amb.unizar.es/people/elena-lanchares/)

#### Dr. Ahmed Fawzy Yousef

Professor, Department Water Resources and Environment International Senior Expert of Project Management And Department of Geology, Desert Research Center, Egypt Email: Ahmedfawzy63@yahoo.com, ahmedfawzy1963@yahoo.com Profile link: NA

#### Dr. Vyacheslav O Vakhnenko

Professor, Division of Geodynamics of Explosion, Subbotin Institute of Geophysics, National Academy of Sciences of Ukrainian, Ukraine Email: vakhnenko@ukr.net

Profile link: NA

#### Dr. A.A. Hanafi-Bojd

Assistant Professor, Department of Medical Entomology & Vector Control, School of Public Health, Tehran University of Medical Sciences, Iran Email: aahanafibojd@tums.ac.ir, aahanafibojd@yahoo.com Profile link: http://tums.ac.ir/faculties/aahanafibojd) Professor,

China University of Petroleum, China Email: cysun@cup.edu.cn Profile link: NA

#### Dr. Ahmed Mohamed El-Waziry

Professor,

King Saud University, College of Food and Agriculture Sciences, Kingdom of Saudi Arabia

Email: aelwaziry@yahoo.com

Profile link: http://faculty.ksu.edu.sa/75830/default.aspx (http://faculty.ksu.edu.sa/75830/default.aspx)

#### Dr. Manoj Gupta

Associate Professor,

Department of Mechanical Engineering, NUS, 9 Engineering Drive 1, Singapore

Email: mpegm@nus.edu.sg

Profile link: http://me.nus.edu.sg/about-us/people/academic-staff/materials/ (http://me.nus.edu.sg/about-us/people/academic-staff/materials/)

#### Dr. Rui Xiao

Professor,

School of Energy and Environment, Southeast University, China

Email: ruixiao@seu.edu.cn

Profile link: http://power.seu.edu.cn/ny\_en/ad/de/c11777a110046/page.htm (http://power.seu.edu.cn/ny\_en/ad/de/c11777a110046/page.htm)

#### Dr. Ming-Chih Shih

Professor,

Department of Health and Nutrition Science,

Chinese Culture University, Taiwan

Email: smz2@faculty.pccu.edu.tw

Profile link: NA

#### Dr. Giovanni Bucci

Professor,

Department of Industrial Engineering and Information and economy,

University of L'Aquila, Italy

Email: giovanni.bucci@univaq.it, bucci@ieee.org

Profile link: https://goo.gl/f6hbDM (https://goo.gl/f6hbDM)

#### Dr. Grzegorz Golanski

Professor, Institute of Materials Engineering, Czestochowa University of Technology, Poland Email: grzegorz.golanski@pcz.pl Profile link: NA

#### Dr. Essam E Khalil

Professor of Energy, Cairo University, Cairo, Egypt Email: khalile1@asme.org Profile link: NA

#### Dr. Santiago Silvestre

Associate Professor, Telecommunications Engineering, Universitat Politècnica de Cataluny, Spain Email: santiago.silvestre@upc.edu Profile link: https://goo.gl/9XaZLz (https://goo.gl/9XaZLz)

#### Dr. Aleksey Aleksandrovich Hlopitskiy

Professor, Department of Technology inorganic substances, Ukrainian State University of Chemical Technology, Ukraine Email: ahlopitskiy@gmail.com Profile link: NA

#### Dr. João Miguel Dias

Associate Professor, Habilitation in Department of Physics, CESAM, University of Aveiro, Portugal Email: joao.dias@ua.pt Profile link: https://goo.gl/pzxoEw (https://goo.gl/pzxoEw)

#### Dr. Jakub Kostecki

Assistant Professor, Faculty of Civil Engineering, Architecture and Environmental Engineering, University of Zielona Góra, Poland Email: j.kostecki@iis.uz.zgora.pl Profile link: NA

#### Dr. Wen Shyang Chow

Professor, School of Materials and Mineral Resources Engineering, Engineering Campus, Universiti Sains Malaysia, Malaysia Email: shyang@usm.my Profile link: https://goo.gl/1sRq6o (https://goo.gl/1sRq6o)

#### Dr. Xu Jianhua

Professor,

Department of Geography, East China Normal University, China

Email: jhxu@geo.ecnu.edu.cn, jhxuecnu@gmail.com

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#### **Dr. Cheng Siong Chin**

Professor,

School of Marine Science and Technology, Newcastle University, UK

Email: cheng.chin@newcastle.ac.uk, cheng.chin@ncl.ac.uk, mcschin1@yahoo.com

Profile link: https://goo.gl/sYFvge (https://goo.gl/sYFvge)

#### **Dr. Vitaly Kober**

Professor, Department of Computer Science, CICESE, Mexico Email: vkober@cicese.mx Profile link: NA

#### Dr. Hamid El Bilali

Centre for Development Research, University of Natural Resources and Life Sciences (BOKU), Vienna, Austria Email: hamid.elbilali@boku.ac.at, elbilali@iamb.it Profile Link: NA

#### Dr. Qing-Wen Wang

Professor,

Department of Mathematics, Shanghai University, P.R. China

Email: wqw@shu.edu.cn

Profile link: http://math.shu.edu.cn/teacher/wangqingwen/ (http://math.shu.edu.cn/teacher/wangqingwen/)

#### Dr. Yu Hai-Liang

Professor, College of Mechanical, and Electrical Engineering, Central South University, China Email: hailiang@uow.edu.au Profile link: https://goo.gl/aRyT2x (https://goo.gl/aRyT2x)

#### Dr. Fernando Reboredo

Professor,

Department of Earth Sciences, New University of Lisbon, Portugal

Email: fhr@fct.unl.pt

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#### Dr. David Coman

Professor,

Medical Director of Paediatrics, The Wesley Hospital, Brisbane, Australia And The Lady Cilento Children's Hospital, Brisbane, Australia

Email: David\_Coman@health.qld.gov.au, david.coman@hotmail.com

Profile link: NA

#### Dr. Sylwia Myszograj

Professor,

Department of Water Technology, Sewage and Wastes,

University of Zielona Gora, Poland Email: S.Myszograj@iis.uz.zgora.pl Profile link: NA

#### Dr. Orlando Manuel da Costa Gomes

Professor of Economics, Lisbon Accounting and Business School (ISCAL), Lisbon Polytechnic Institute, Portugal Email: omgomes@iscal.ipl.pt, Profile link: NA

#### Dr. Kleopatra Nikolopoulou

Professor, School of Education, University of Athens, Athens, Greece Email: klnikolop9@yahoo.gr Profile link: NA

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Professor,

Department of Computer Science and Engineering,

University of Calcutta, India

Email: 1954samir@gmail.com

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Professor,

Solid and Structural Mechanics,

University Mediterranea of Reggio Calabria, Italy

Email: aurora.pisano@unirc.it

Profile link: https://www.unirc.it/scheda\_persona.php?id=680 (https://www.unirc.it/scheda\_persona.php? id=680)

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Professor,

College of Agricultural Sciences,

International University of Business Agriculture and Technology (IUBAT University), Bangladesh

Email: farjana1s@iubat.edu, farjana1s@ymail.com

Profile link: https://iubat.edu/web1/index.php/team-details/dr-farjana-sultana/ (https://iubat.edu/web1/index.php/team-details/dr-farjana-sultana/)

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Professor,

Department of Agricultural Economics, Faculty of Agriculture, Ege University, Turkey

Email: sait.engindeniz@ege.edu.tr, saitengindeniz@gmail.com

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Professor, Department of Agrochemistry and Environment, University Miguel Hernandez, Spain Email: jonavar@umh.es Profile Link: NA

#### Dr. Gregory J. Grigoropoulos

Professor, Ship and Marine Hydrodynamics at the School of Naval Architecture and Marine Engineering (SNAME), National Technical University of Athens (NTUA), Greece Email: gregory@central.ntua.gr, gregory@mail.ntua.gr Profile Link: NA <u>Short Biosketch</u> (http://www.journalcjast.com/index.php/CJAST/gregory-j-grigoropoulos)

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Senior Lecturer, Head of Chemical and Polymer Innovation Technology Cluster, Faculty of Chemical Engineering Technology, TATI University College, Jalan Panchor, Terengganu, Malaysia Email: drahmedtati@gmail.com Profile Link: http://tatiuc.edu.my/ftkk/penerbitan.html (http://tatiuc.edu.my/ftkk/penerbitan.html) Short Biosketch (http://www.journalcjast.com/index.php/CJAST/ahmed-h-a-dabwan)

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Professor,

Instituto de Química, Universidad Nacional Autónoma de México,

Circuito Exterior S/N. Ciudad Universitaria. Coyoacán, México

Email: damor@unam.mx

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#### Dr. Rahul Prabhakar More

Professor, Abeda Inamdar Senior College Of Arts, Science & Commerce, Azam Campus, Camp, Pune, India Email: rahul\_more25@yahoo.com, morerahul.25@gmail.com Profile Link: NA <u>Short Biosketch</u> (http://www.journalcjast.com/index.php/CJAST/rahul-prabhakar-more)

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Professor, Department of Mathematics, COMSATS Institute of Information Technology Attock Campus, Pakistan Email: numantng@gmail.com Profile Link: NA

#### Dr. Abdullah Aydin

Associate Professor, Department of Science Education, Kırşehir Ahi Evran University, Turkey Email: aaydin@ahievran.edu.tr, aydinch@gmail.com

Profile Link: https://akademik.ahievran.edu.tr/site/abdullahaydin (https://akademik.ahievran.edu.tr/site/abdullahaydin)

#### Dr. Maduike Chiehiura Onwubiko Ezeibe

Professor, Department of Veterinary Medicine, Michael Okpara University of Agriculture, Umudike, Abia State, Nigeria Email: maduikeezeibe@yahoo.com Profile Link: NA <u>Short Biosketch</u> (http://www.journalcjast.com/index.php/CJAST/maduike-chiehiura-onwubiko-ezeibe)

#### Dr. Belisario Dominguez-Mancera

Laboratory of Cell Biology and Radioimmunoassay, School of Veterinary Medicine and Animal Science, University Veracruz, Mexico Email: beldominguez@uv.mx Profile Link: https://www.uv.mx/personal/beldominguez (https://www.uv.mx/personal/beldominguez)

#### Dr. Seham Mohammed Elkassas

Department of Animal Wealth Development, Faculty of Veterinary Medicine, Kafrelsheikh University, Kafrelsheikh, Egypt. Email: seham.elkassas@vet.kfs.edu.eg, seham618@yahoo.com Profile Link: NA <u>Short Biosketch</u> (http://www.journalcjast.com/index.php/CJAST/seham-mohammed-elkassas)

#### Dr. Azza Hashim Abbas Babikir

Department of Petroleum Engineering, Asia Pacific University, Bangladesh Email: azzahashim2008@gmail.com,haazza2@utm.live.my Profile Link: NA

#### Dr. Pavel K. Kepezhinskas

PNK GeoScience, Tampa, Florida, USA

Email: pavel\_k7@yahoo.com Profile Link: NA

#### Dr. Umar Nirmal

Senior Lecturer, Centre of Advanced Mechanical and Green Technology, Faculty of Engineering and Technology, Multimedia University, Malaysia Email: nirmal@mmu.edu.my, nirmal288@zoho.com Profile Link: https://mmuexpert.mmu.edu.my/nirmal (https://mmuexpert.mmu.edu.my/nirmal)

#### Dr. Nagesh Peddada

Department of Biophysics, University of Texas Southwestern Medical Center, USA Email: nagesh.peddada@utsouthwestern.edu, nagesh.mku@gmail.com Profile Link: NA

#### Dr. Ana Pedro

Professor,

Department of Education,

University of Aveiro, Aveiro, Portugal

Email: ana.pedro@ua.pt

Profile Link: NA

Dr. Md. Hossain Ali

Chief Scientific Officer, and Head,

Agril. Engg. Division,

Bangladesh Institute of Nuclear Agriculture (BINA)

Bangladesh Agricultural University Campus,

Bangladesh

Email: mh\_ali@bina.gov.bd (mailto:mh\_ali@bina.gov.bd), hossain.ali.bina@gmail.com (mailto:hossain.ali.bina@gmail.com),

mha\_bina@yahoo.com (mailto:mha\_bina@yahoo.com)

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#### **Dr. Diony Alves Reis**

Federal University of the West of Bahia, Brazil Email: dionyodin@gmail.com, diony.reis@ufob.edu.br Profile Link: NA

#### Dr. Oner Cetin

Professor, Department of Irrigation Engineering, Agricultural Faculty, Dicle University Diyarbakir, Turkey Email: oner\_cetin@yahoo.com, onercetin@dicle.edu.tr Profile Link: https://goo.gl/cgtbT3 (https://goo.gl/cgtbT3)

#### Dr. Ogunlade, Clement Adesoji

Lecturer, Department of Agricultural Engineering, Faculty of Engineering, Adeleke University, Nigeria Email: clement2k5@yahoo.com, clementlawmaker@gmail.com, ogunlade.clement@adelekeuniversity.edu.ng Profile Link: NA

#### Dr. P. Senguttuvel

Indian Institute of Rice Research (Formerly Directorate of Rice Research), India Email: Senguttuvel@gmail.com, P.Senguttuvel@icar.gov.in, velusen09@gmail.com Profile Link: NA

#### **Dr. Ashish Anand**

Department of Orthopaedic Surgery, GV Montgomery Veteran Affairs Medical Center, USA Email: ashishanandortho@yahoo.com Profile Link: NA

#### Dr. Tushar Ranjan

Assistant Professor, Department of Molecular Biology & Genetic Engineering, Bihar Agricultural University, Sabour, India Email: mail2tusharranjan@gmail.com Profile Link: NA

#### Dr. Bishun Deo Prasad

Assistant Professor, Department of Molecular Biology & Genetic Engineering, Bihar Agricultural College, Bihar Agricultural University, India Email: bdprasadbau@gmail.com Profile Link: NA

#### Dr. Sangita Sahni

Assistant Professor, Department of Plant Pathology, Tirhut College of Agriculture, Dr. Rajendra Prasad Central Agricultural University, India Email: sangitampp@gmail.com Profile Link: NA

#### Dr. Awadhesh Kumar Pal

Assistant Professor, Department of Biochemistry and Crop Physiology, Bihar Agricultural University, India Email: awapal@gmail.com Profile Link: NA

#### Dr. Aydin Unay

Professor, Department of Field Crops, Faculty of Agriculture, University of Aydin Adnan Menderes, Turkey Email: aunay@adu.edu.tr Profile Link: NA

#### Dr. Ekpenyong, Christopher Edet

Associate Professor, College of Health Sciences, University of Uyo, Nigeria Email: chrisvon200@yahoo.com Profile Link: NA

#### Dr. Abida Farooqi

Assistant Professor,

Department of Environmental Sciences, Quiad-i-Azam University, Pakistan

Email: missfarooqi@hotmail.com

Profile link: https://www.qau.edu.pk/profile.php?id=931002 (https://www.qau.edu.pk/profile.php? id=931002)

#### Dr. Stănilă Andreea

Profesor

Department of Food Science and Technology,

University of Agricultural Sciences and Veterinary Medicine, Cluj-Napoca, Romania

Email: andreea.stanila@usamvcluj.ro

Profile Link: NA

#### Dr. Orujova Naila Hidayat

Professor Department of soil science and agricultural sciences Institute of Soil Science and Agricultural Chemistry, National Academy of Sciences of Azerbaijan, Azerbaijan Email: naila.56@mail.ru Profile Link: NA

#### Dr. Rajan Bhatt

Department of Soil Science Punjab agricultural University, Ludhiana, Punjab, India Email: rajansoils@pau.edu, rajansoils@pau.edu Profile Link: NA

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## Effect of Temperature and Long Storage of Fatty Acid Profile, Value Peroxide and Value TBA Fillet of Snapper (Lutjanus sp)

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#### **Rahim Husain**

Faculty of Fisheries and Marine Science, Gorontalo State University, Jend. Sudirman, No. 6, Gorontalo City, Indonesia.

#### Rieny Sulistijowati

Faculty of Fisheries and Marine Science, Gorontalo State University, Jend. Sudirman, No. 6, Gorontalo City, Indonesia.

## Abstract

This research was conducted at the Fisheries Technology Laboratorium Department of Fisheries and Marine Sciences, Gorontalo State University, at the Mey 2018 for three months in a month. Fish has a high nutritional value and is a major food source in many countries. Lipid fish has a high content of polyunsaturated acids (PUFA), especially eicosapentaenoic acid (EPA;20:5n-3) and docosahexenoate acid (DHA; 22: 6n-3). This study aims to the impact effect of temperature and storage time of the fatty acid snapper fillet (*Lutjanus sp*) and damage caused by the storage process. The sample selection stage is Snapper (*Lutjanus sp*) fresh weeds, and the meat is released by gills, bones, then washed and rinsed with ice water to remove blood and other debris. Contineus does the cleavage from head to tail without causing his back to be disturbed. Filleting is done by slicing the meat ribs longitudinally to produce boneless meat. Stored at temperatures of 0, 10, 20, 30 and 40°C for 45 days. The results of fatty acid analysis snapper fillet (*Lutjanus sp*) has a saturated fatty acid lauric acid, acid tridecanoic, myristate acid, pentadecanoic acid, palmitic acid, stearic acid, heneicosanoic acid, behenic acid, lignoceric acid, acid and acid heptadecanoate arachidic. Monounsaturated fatty acids oleic acid, nervonic acid, palmitoleic acid, acid and acid erucat eicosanoic, while polyunsaturated fatty acids eicosatetraenoate acid, dokosadinoat

acid, eicosapentaenoic acid, eikosatrioanat acid, arachidonic acid and linolenic acid are susceptible to oxidative damage. Increased primary peroxide snapper fillet (*Lutjanus sp*) the higher the storage temperature increases from 0°C to 40°C. TBA value increases from 0° to 40°C with increased storage time of one day to 45 days storage time. From the results of this study found a way for Snapper fish fillets (*Lutjanus sp*) to have a longer expiration in storage, from 1 day to 45 days without reducing the Omega and DHA content.

Keywords:

Fillet snapper (Lutjanus sp), fatty acids, peroxide value and TBA value.

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### Effect of Temperature and Long Storage of Fatty Acid Profile, Value Peroxide and Value TBA Fillet of Snapper (*Lutjanus sp*)

#### Rahim Husain<sup>1\*</sup> and Rieny Sulistijowati<sup>1</sup>

<sup>1</sup>Faculty of Fisheries and Marine Science, Gorontalo State University, Jend. Sudirman, No. 6, Gorontalo City, Indonesia.

#### Authors' contributions

This work was carried out in collaboration between both authors. Both authors read and approved the final manuscript.

#### Article Information

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#### ABSTRACT

This research was conducted at the Fisheries Technology Laboratorium Department of Fisheries and Marine Sciences, Gorontalo State University, at the Mey 2018 for three months in a month. Fish has a high nutritional value and is a major food source in many countries. Lipid fish has a high content of polyunsaturated acids (PUFA), especially eicosapentaenoic acid (EPA;20:5n-3) and docosahexenoate acid (DHA; 22: 6n-3). This study aims to the impact effect of temperature and storage time of the fatty acid snapper fillet (*Lutjanus sp*) and damage caused by the storage process. The sample selection stage is Snapper (*Lutjanus sp*) fresh weeds, and the meat is released by gills, bones, then washed and rinsed with ice water to remove blood and other debris. Contineus does the cleavage from head to tail without causing his back to be disturbed. Filleting is done by slicing the meat ribs longitudinally to produce boneless meat. Stored at temperatures of 0, 10, 20, 30 and 40°C for 45 days. The results of fatty acid analysis snapper fillet (*Lutjanus sp*) has a saturated fatty acid lauric acid, acid tridecanoic, myristate acid, pentadecanoic acid, palmitic acid, stearic acid, heneicosanoic acid, behenic acid, lignoceric acid, acid and acid heptadecanoate

arachidic. Monounsaturated fatty acids oleic acid, nervonic acid, palmitoleic acid, acid and acid erucat eicosanoic, while polyunsaturated fatty acids eicosatetraenoate acid, dokosadinoat acid, eicosapentaenoic acid, eikosatrioanat acid, arachidonic acid and linolenic acid are susceptible to oxidative damage. Increased primary peroxide snapper fillet (*Lutjanus sp*) the higher the storage temperature increases from 0°C to 40°C. TBA value increases from 0° to 40°C with increased storage time of one day to 45 days storage time. From the results of this study found a way for Snapper fish fillets (*Lutjanus sp*) to have a longer expiration in storage, from 1 day to 45 days without reducing the Omega and DHA content.

Keywords: Fillet snapper (Lutjanus sp); fatty acids; peroxide value and TBA value.

#### **1. INTRODUCTION**

In general, the fish has no nutritional value and is a major food source in many countries. Lipid fish has a high content of polyunsaturated acids (PUFA), especially eicosapentaenoic acid {EPA; 20: 5n-3} and docosahexenoate acid (DHA; 22: 6n-3) [1,2]. Processing by freezing the fish has been used for thousands of years because of the quality and high product [3]. The concept of storage by freezing depends on the product temperature decrease to slow decay so that when the fish melted freshness can be maintained [4].

However, fish and fishery products may undergo undesirable changes during storage and can damage the storage time limit. Unwanted changes resulting from the oxidation of proteins [5,6] and lipid oxidation [7,8]. Fish protein experienced a number of changes (cause cannot form aggregates) that modify the structural and functional properties of fish muscle [9].

Degradation of polyunsaturated fatty acids (PUFA) by lipid oxidation during storage led to the formation of volatiles associated with rancidity [1]. The high level of unsaturated fats makes the fish tissue is very susceptible to peroxidation and easily damaged. Oxidative changes primarily related to the taste and texture of the fish. In later stages of the process of lipid peroxidation, changes in color and nutritional value will be observed or secondary products of lipid [10]. This study aims to determine the effect of temperature and storage time of the fatty acid snapper fillet (*Lutjanus sp*) and damages resulting from the storage process.

#### 2. MATERIALS AND METHODS

#### 2.1 Raw Material

Raw materials snapper (*Lutjanus sp*) obtained from the fish auction place (TPI) kobong, Village Kaligawe, Semarang, Central Java.

#### 2.2 Sample Preparation Fish Fillets

The sample selection stage is Snapper (*Lutjanus* sp) fresh weeds, and the meat is released by gills, bones, then washed and rinsed with ice water to remove blood and other debris. Contineus does the cleavage from head to tail without causing his back to be disturbed. Filleting is done by slicing the meat ribs longitudinally to produce boneless meat. Stored at temperatures of 0, 10, 20, 30 and 40°C for 45 days.

#### 2.3 Proximate Analysis

Chemical analysis: Proximate moisture content, ash content, protein and fat by AOAC [11].

#### Fatty Acid Analysis Procedure

#### a. Hydrolysis

After selecting the sample, then the sample is cleaned with water, then mashed and weighed  $\pm$  10 g of the sample. Hydrolysis with 10 ml HCl. then the water is heated at a temperature of 70°C, boiling. Cold. Extract with 15 ml of diethyl ether and 15 ml of oil, take the top layer. Extract again with 15 ml of diethyl ether and 15 ml of oil, take the top layer, make one with the previous results. Steamed in a water bath with the help of N2.

### b. Analysis of saturated fatty acids and unsaturated

Taken 0.5 ml sample, add 1.5 ml of sodium methanolic solution, cover and heat at 70°C for 5-10 minutes while being shaken. Chill. Add 2 ml Boron trifluoride ethanoic, heat at 70°C for 5-10 minutes. Chill. Extract with 1 ml of heptane and 1 ml saturated NaCl. Take the top layer and put in Eppendorf. Injected into the GC. Injected as much 1 $\mu$  samples on GC - 2010 SHIMADZU.

#### 2.4 Analysis Value Peroxide

Samples (0.5 g) put in a test tube, then added 0.1 ml of a solution of ammonium thiocyanate and 0.1 ml of solution feroklorida. The test tube is shaken for 5 seconds and heated at  $50^{\circ}$ C for 2 minutes, then cooled to a temperature of  $25^{\circ}$ C. Use absorbance using a spectrophotometer at a wavelength of 510 nm. A blank solution prepared using all solvent without the sample (Hills and Thiel, 1946 modified Adnan [12]).

#### 2.5 Analysis Value TBA (Expand)

Analysis of figures TBA performed according to the method Tokur and Korkmaz [13]. Oil added 0.5 g in 50 ml of distilled water, then add another 2.5 mL N HCl then distilled. Absorb distilled to 50 ml, 5 ml of distilled download then added with 5 ml TBA. Thereafter, heated for 30 minutes and cooled. Absorbance at a wavelength of 528 nm. Value TBA = mg malonaldehyde / kg of oil.

#### 3. RESULTS AND DISCUSSION

#### 3.1 Proximate Fish Snapper (*Lutjanus sp*)

Based on Table 1 fat content snapper (*Lutjanus* sp) in the study was 1.96% compared to that reported by de Castro et al. [14] on tilapia (*Oreochromis niloticus*), i.e 0.79% and tambagui (*Colossoma macropomum*) i.e 1.30%. The fat content rough on snapper (1.96%) is still higher, it indicates that the snapper (*Lutjanus* sp) belong to the fish that contains high fat enough.

### Table 1. Proximate fish snapper (Lutjanus sp) and Fe content

No	Chemical composition	Amount (%)
1.	Moisture	78,39
2.	Ash	1,58
3.	Protein	18,77
4.	Fat	1,96
5.	Carbohydrate by difference	0,30
6.	Fe (ppm)	121,7
D	escription: Data is derived	from repeat 3x

Differences in fish fat content are strongly influenced by the type of fish, fish size, fishing

season and the environment in which the fish live. According to Shaviklo [15] demersal fish higher in fat than pelagic fish that lives in surface waters. Demersal fish usually live in the bottom waters and rarely engage in the activity. In Table 2 also shows the snapper (*Lutjanus sp*) containing Fe total was 121.47 ppm. Okada report [16] describes the color of the flesh of fish due to the Fe content in the meat is very high because it is rich hemoprotein (80%) mainly myoglobin and hemoglobin. Based Okada [16], the content of white meat hemoprotein red snapper low so the meat is white.

#### 3.2 Composition Fatty Acid

Based on the analysis of fatty acids snapper fillet (Lutjanus sp) are listed in Table 4, snapper fillet (Lutjanus sp) has saturated fatty acids (Saturated Fatty Acid, SFAs), which consists of lauric acid, tridecanoic, acid myristate, acid acid pentadecanoic, palmitic acid, stearic acid, heneicosanoic acid, behenic acid, lignoceric acid, acid and acid heptadecanoatearachidic. Acids monounsaturated fats (MUFA) consisting of: oleic acid, acid nervonat, palmitoleic acid, acid erucat and sour eicosanoic. while the acid unsaturated fatty compound (Polv Unsaturated Fatty Acids, PUFAs) consisting of acid eicosatetraenoate, dokosadinoat acid. eicosapentaenoic acid, eikosatrioanat acid arachidonic acid and linolenic acid are susceptible to oxidative damage.

According Nazemroaya et al. [17]; Karami et al. [18] that when the fish are kept in a long time then, the amount of saturated fatty acids (Saturated Fatty Acid, SFA) will increase as well fatty acids monounsaturated (Mono as Unsaturated Fatty Acid, MUFA) will be slightly experience improved although not too significant. polyunsaturated fatty acids While (Poly Unsaturated Fatty Acids, PUFAs) experienced a reduction process. This is similar to the research conducted at the snapper fillet (Lutjanus sp) during storage.

Aranda et al. [19] states that the oxidation of PUFAs begins with the production of hydrogen peroxide with three different ways: (1) autooksidasi, (2) enzymatic oxidation, and (3) photooxidation, which can also occur simultaneously. Product peroxide decomposes by different mechanisms, forming a secondary oxidation products; both the primary and secondary oxidation products can react with amino groups of proteins, producing compounds that modify the interaction of taste, smell and functional properties of proteins.

Furthermore, the fraction of volatile compounds is indicated as the cause of rancidity in fatty fish [19]. Secbecic and Beutelspecher [20] states that the oxidation is the process of fat breakdown and lead to the formation of off-flavor compounds and condition is called rancid (rancid). Processed food products were rancid, may experience discoloration and loss of nutritional value for the oxidation of unsaturated fatty acids (PUFA) that impact on the quality. The compound oxide such as peroxides, aldehydes, and ketones harmful to human health. While McClement and Decker [21] states the factors that affect the speed of oxidation include the number and type of oxygen, the chemical structure of lipids, antioxidant and prooxidant form: ferrous metals, sensitiser, storage temperature and properties of packaging materials.

## 3.3 Effect of Temperature and Time against Value Peroxide

Value peroxide as primary products of oxidation snapper fillet (*Lutjanus sp*) during storage at different temperatures and times can be seen in Table 3.

In the treatment of frozen storage temperature (0°C) shows the formation of peroxides to 45 days of storage. Speed peroxide formation increased 9.6 times at a temperature of 0°C with a storage temperature increases, while at a temperature of 10, 20, 30, and 40°C peroxide value increased respectively 14.33, 16.12, 19.19, and 23.05 times meq/kg sample with increased storage temperature. The peroxide values increased from 0° to 40°C with increased storage time of one day to 45 days storage time. Increased primary peroxide snapper fillet (*Lutjanus sp*) the higher the storage temperature increases from 0°C to 40°C.

No	Storage temperature (°C)					
	Profil asam lemak (%)	0°C	10°C	20°C	30°C	40°C
1	Lauric acid	0,703	-	-	0,99	-
2	Tridecanoac acid	0,001	-	-	0,86	-
3	Myristic acid	4,687	0,74	4,44	2,69	0,51
4	Pentadecanoac acid	0,001	0,90	1,03	2,46	0,38
5	Palmetic acid	2,283	1,67	4,39	3,58	12,94
6	Stearic acid	0,708	-	9,19	15,46	12,14
7	Heneicosanoac acid	13,462	-	0,78	2,03	-
8	Behenac acid	-	1,04	0,65	0,68	1,54
9	Lignoserat acid	0,762	-	0,74	-	0,55
10	Heptadecanoac acid	17,811	-	1,96	-	-
11	Arachidac acid	2,945	-	-	-	-
I	Saturated Fatty Acid (SFA)	25,552	4,35	32,73	31,75	28,06
1	Oleic acid	8,807	12,63	16,37	22,70	10,82
2	Nervonac acid	1,64	1,34	1,03	1,27	-
3	Palmitoleic acid	13,239	2,59	-	-	-
4	Erucat acid	-	5,43	5,26	5,43	3,58
5	Eicosanoac acid	0,001	8,34	-	-	-
11	Mono Unsaturated Fatty Acid(MUFA)	23.687	23,72	22,66	29,4	14,4
1	Eicosatetraenoic acid	-		-	5,60	1,05
2	Docosadinoac acid	0,827	-	0,43	-	0,70
3	Eicosapentanoac acid	6,055	3,52	2,29	0,43	-
4	Eicosatrienoac acid	1,557	2,6,23	9,82	-	-
5	Arachidonac acid	0,692	-	-	-	-
6	Docosaheksanoac acid	5,31	-	0,17	0,25	-
7	Linoleict acid	10, 612	1,52	3,27	2,56	3,57
	Poly Unsaturated Fatty Acid (PUFA)	25,055	14,99	15,98	8,84	10,32
Total		74,294	103.07	101.37	99,99	82,78

Table 2. Composition f	atty acids of snapper fillet (	<i>Lutjanus sp</i> ) during storage

Description: Data is derived from repeat 2x

	Temper	ature 0°C
N0	Storage temperature (day)	Peroxide Value (meq/kg)
1	0	1.7411
2	5	2.9437
3	10	4.06055
4	15	5.296
5	20	6.7797
6	25	6.29125
7	30	10.905
8	35	11.4923
9	40	12.98225
10	45	13.21835
	Tempera	ture 10°C
N0	Storage temperature (day)	Peroxide Value (meq/kg)
1	0	1.7411
2	3	3.26525
3	6	5.63635
4	9	6.90375
5	12	6.3911
6	15	9.60175
7	18	10.78685
8	21	10.4857
9	24	14.5174
10	27	15.9977
	Tempera	ture 20°C
N0	Storage temperature (day)	Peroxide Value (meq/kg)
1	0	1.7411
2	1	4.21665
3	2	6.96335
4	3	8.3699
5	4	8.5134
6	5	11.22005
7	6	11.963
8	7	15.59645
9	8	15.9638
10	9	18.0701
	Tempera	ture 30°C
N0	Storage temperature (day)	Peroxide Value (meq/kg)
1	0	1.7411
2	0.5	4.5663
3	1	7.06935
4	1.5	9.2808
5	2	10.422
6	2.5	12.1345
	3	11.83165
7	5	
	3.5	16.974
7 8 9		

#### Table 3. Date analysis peroxide value snapper fillet (*Lutjanus sp*)

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Temperature 40°C			
N0	Storage temperature (day)	Peroxide Value (meq/kg)	
1	0	1.7411	
2	0.25	4.67575	
3	0.5	7.7808	
4	0.75	10.7164	
5	1	12.06535	
6	1.25	12.6637	
7	1.5	18.4146	
8	1.75	18.42355	
9	2	22.9266	
10	2.25	24.62795	

Description: Data is derived from repeat 3x

Table 4. Data analysis value TBA snapper fillet (Lutjanus sp)
---

	Temp	perature 0°C
No	Storage temperature (day)	Value TBA (mg MDA/kg)
1	0	2.307378
2	5	3.504316
3	10	5.277034
4	15	6.573477
5	20	6.315832
6	25	9.565049
7	30	11.91097
8	35	11.75207
9	40	13.71696
10	45	14.62961
		erature 10°C
No	Storage temperature (day)	Value TBA (mg MDA/kg)
1	0	2.307378
2	3	4.299369
3	6	6.748229
4	9	6.105547
5	12	10.76352
6	15	13.41159
7	18	13.2912
8	21	15.61061
9	24	16.80039
10	27	17.21704
	Temp	erature 20°C
No	Storage temperature (day)	Value TBA (mg MDA/kg)
1	0	2.307378
2	1	4.424119
3	2	6.195105
4	3	6.932098
5	4	10.79865
6	5	13.33775
7	6	13.45672
8	7	17.74074
9	8	18.62311
10	9	20.31495

Temperature 30°C					
Storage temperature (day)	Value TBA (mg MDA/kg)				
0	2.307378				
0.5	5.887911				
1	7.589422				
1.5	8.931236				
2	8.773049				
2.5	13.85214				
	14.5131				
3.5	14.5922				
4	20.7325				
4.5	22.03099				
Temperature 40°C					
Storage temperature (day)	Value TBA (mg MDA/kg)				
0	2.307378				
0.25	5.975746				
0.5	7.946435				
0.75	8.185896				
1	8.54802				
1.25	10.79128				
-					
_					
	Storage temperature (day)           0           0.5           1           1.5           2           2.5           3           3.5           4           4.5           Temp           Storage temperature (day)           0           0.25           0.5           0.75           1	Storage temperature (day)         Value TBA (mg MDA/kg)           0         2.307378           0.5         5.887911           1         7.589422           1.5         8.931236           2         8.773049           2.5         13.85214           3         14.5131           3.5         14.5922           4         20.7325           4.5         22.03099           Temperature 40°C           Storage temperature (day)         Value TBA (mg MDA/kg)           0         2.307378           0.25         5.975746           0.5         7.946435           0.75         8.185896           1         8.54802           1.25         10.79128           1.5         15.43709           1.75         15.59018           2         20.52694			

Description: Data is derived from repeat 3x

According to Pak et al. [22], peroxide value is an indicator of the stability of the oil against oxidation, the oxidation products of primary lipid parameters, namely hydroperoxide. Oxidation of lipids/oils naturally easily occurs, because tuna fish oil rich in PUFAs (6 double bonds), while oil containing many double bonds susceptible to lipid oxidation reactions. Thus, the oxygen molecules that are attached to the double bond susceptible to oxidation.

#### 3.4 Effect of Temperature to Value TBA

Value TBA is used to measure the secondary products of lipid oxidation, especially coming from PUFA (hide no, 2012) and indicates the level of rancidity, especially in the high PUFA-containing oil [23]. TBA formation as a secondary oxidation products snapper fillet (*Lutjanus sp*) during storage can be seen in Table 4.

Value TBA increased 7.21 times at a temperature of 0°C with an increase in storage temperature, whereas at a temperature of 10, 20, 30, and 40°C peroxide value increased respectively to 9.19, 9.89, 10.63, and 12.59 times the MDA mg/kg of the sample with increased storage temperature. Value TBA increases from 0° to 40° with increased storage time of one day to 45 days storage time. In the treatment of

storage temperature of 0°C-40°C showed the formation of value TBA during storage [23].

#### 4. CONCLUSION

Differences in fat content of the fish are strongly influenced by the freshness of the fish is used as a critical factor in resulted isolates, methods of isolation/extraction is used, homogenization of meat processing, the ratio of fish and solvents (viscosity) used, length of extraction, time and temperature processing and dissolution protein. Snapper fillet (Lutianus sp) has a saturated fatty acid comprising: lauric acid, acid tridecanoic, myristate acid, pentadecanoic acid, palmitic acid, stearic acid, heneicosanoic acid, behenic acid, acid lignoceric acid. and acid heptadecanoatearachidic. Fatty acids are monounsaturated consisting of: oleic acid, acid nervonat, palmitoleic acid, acid erucat and sour eikosanoat, while the unsaturated fatty acid compound consisting of: acid eicosatetraenoate, acid dokosadinoat acid, eicosapentaenoic acid eikosatrioanat, arachidonic acid and linolenic acid susceptible to oxidative damage.

Speed formation of peroxide value increased 9.6 times at a temperature of 0°C with a storage temperature increases, while at a temperature of 10, 20, 30, and 40°C peroxide value increased

respectively 14.33, 16.12, 19.19 and 23.05 times meq/kg sample with increased storage temperature. Value TBA increased 7.21 times at a temperature of 0°C with an increase in storage temperature, whereas at a temperature of 10, 20, 30, and [10] 40°C peroxide value increased respectively to 9.19, 9.89, 10.63, and 12.59 times the MDA mg/kg of the sample with increased storage temperature.

#### **COMPETING INTERESTS**

Authors have declared that no competing interests exist.

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